



Review Article

Flavin-containing Monooxygenases in the Brain and their Involvement in Neurodegeneration and Aging



Boyu Li and Zhuoling An*

Department of Pharmacy, Beijing Chao-Yang Hospital, Capital Medical University, Beijing, China

Received: August 15, 2022 | Revised: December 27, 2022 | Accepted: March 17, 2023 | Published online: April 28, 2023

Abstract

Flavin-containing monooxygenases (FMOs) catalyze the oxygenation of a diverse range of sulfur or nitrogen-containing xenobiotics. Recently accumulated evidence has demonstrated the roles of FMOs in physiological and pathological conditions, including neurodegeneration and aging. However, the mechanisms underlying their functions are poorly understood. In this review, we summarize the expression and localization of FMOs in the brain, the endogenous chemicals and xenobiotics metabolized by FMOs, and the consequences of FMO deficiency. The understanding of FMOs activity in the brain is important for fully elucidating the roles of FMOs in pathological mechanisms.

Introduction

Flavin-containing monooxygenases (FMOs) constitute a family of microsomal enzymes catalyzing the oxidation of nucleophilic heteroatom-containing xenobiotics.¹ They oxygenate the sulfur or nitrogen atoms in chemicals with soft nucleophiles.² FMOs are involved in the pathogenic process of trimethylaminuria, atherosclerosis, cardiovascular disease, diabetes, and metabolic disorders.^{3–6} In recent years, the involvement of FMOs in neurodegeneration and aging has emerged,⁷ but the underlying mechanisms have not been elucidated. In this review, we summarize the expression and localization of FMOs in the brain, the endogenous chemicals and xenobiotics metabolized by FMOs in the brain, and the consequences of FMO deficiency.

FMO

FMO (EC 1.14.13.8) was first described by Ziegler *et al.*^{8,9} Humans possess five functional FMO genes, designated *FMO1–5*. *FMO1–4* are clustered on chromosome 1 q24.3, and *FMO5* is lo-

cated at 1q21.1.^{10,11} Numerous allelic variants, including approximately 20 of human *FMO1*, have been reported.¹²

Mammalian FMOs are NADPH- and oxygen-dependent microsomal monooxygenases that usually metabolize nitrogen- and sulfur-containing compounds.^{1,13,14} The catalytic mechanism involves a first step in which FAD undergoes a 2-electron reduction by NADPH. The reduced flavin then reacts rapidly with molecular oxygen to form peroxyflavin. This nucleophilic attack by the substrate on FADOOH results in the transfer of one atom of molecular oxygen to the substrate with another contributing to the formation of water.

Trimethylaminuria is a currently confirmed rare inherited metabolic disorder associated with abnormal amounts of dietary-derived trimethylamine and is caused by the mutations in *FMO3*.^{15,16}

Emerging roles of FMOs in neurodegeneration and aging

Amyotrophic lateral sclerosis

Association between FMOs and amyotrophic lateral sclerosis (ALS) has been widely reported although some reports are contradictory. Malaspina *et al.*¹⁷ reported an 80% reduction in *FMO1* mRNA levels in the spinal cord of sporadic ALS patients. In contrast, Gagliardi *et al.*¹⁸ observed greater *FMO1* expression in the spinal cord and brain stem of ALS patients compared with that in healthy controls. Gagliardi *et al.*¹⁹ found that the mRNA levels of all FMOs except for *FMO3* were up-regulated in the brain of SOD1-mutated (G93A) ALS mice compared with control mice, with the highest increase in *FMO1* in the spinal cord and brainstem. Cereda *et al.*¹² found a significantly elevated frequency of *FMO1* single nucleotide polymorphisms in female sporadic ALS patients, further indicating that specific allelic variants of *FMO1* might be associated with ALS development.

Keywords: Flavin-containing monooxygenases (FMOs); Brain; Expression; Location; Substrates; Deficiency; Neurodegeneration and aging.

Abbreviations: ALS, amyotrophic lateral sclerosis; CYP, cytochrome P450; FMOs, flavin-containing monooxygenases; MPP⁺, 1-methyl-4-phenylpyridinium; MPTP, 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine; PD, Parkinson's disease; PTP, 4-phenyl-1,2,3,6-tetrahydropyridine; TMAO, trimethylamine N-oxide; NADPH, nicotinamide adenine dinucleotide phosphate oxidase; FAD, flavin adenine dinucleotide.

***Correspondence to:** Zhuoling An, Department of Pharmacy, Beijing Chao-yang Hospital, Capital Medical University, Beijing 100020, China. ORCID: <https://orcid.org/0000-0002-7996-5002>. Tel: +86 010 85231362, Fax: +86 010-8523-1362, E-mail: anzhuoling@163.com

How to cite this article: Li B, An Z. Flavin-containing Monooxygenases in the Brain and their Involvement in Neurodegeneration and Aging. *J Explor Res Pharmacol* 2023;8(3):237–241. doi: 10.14218/JERP.2022.00067.

Parkinsonism

Accumulating evidence indicates a relationship between FMOs and parkinsonism. The FMO gene cluster is associated with the volume of the lentiform nucleus, which is a physiological marker associated with Parkinson's disease (PD). Nicotine can be N-oxidized by FMOs and can reduce oxidative stress and neuro-inflammation in the brain and improve synaptic plasticity and neuronal survival of dopaminergic (DA) neurons, thereby benefiting PD patients.^{20,21} MPTP (1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine) is a neurotoxin and its toxic metabolite 1-methyl-4-phenylpyridinium (MPP⁺) can kill DA neurons and elicit parkinsonism. MPTP can be deactivated by FMOs into a harmless metabolite in the brain (discussed in detail in the section, *Endogenous Substances and Xenobiotics Oxygenated by FMOs in the Brain*). In addition, we have shown that FMO1 deficiency promotes neuroinflammation that affects the survival of DA neurons in mice. The levels of *FMO1* mRNA transcripts decreased in a rotenone model of parkinsonism, accompanied by decreasing levels of *parkin* mRNA transcripts and increased Caspase-3 activation.^{22,23}

Aging

FMO1–5 have all been reported to be transcriptionally activated in classical mouse models of longevity, including calorie restriction, growth hormone/insulin-like growth factor 1 signaling disruption, and rapamycin treatment.⁷ The expression of *FMO3* is up-regulated in the liver of a variety of longevity mouse models.^{24–27} However, up-regulation of *FMO3* expression in hepatocytes of murine models has recently been shown to prevent or reverse hepatic aging. This mimicked calorie restriction and the associated mechanism is probably attributed to the promotion of autophagy.²⁸ Furthermore, feeding with a normal diet significantly down-regulated *FMO1* mRNA transcripts in mice in an age-dependent manner,²⁹ indicating that reduced *FMO1* expression contributes to the progression of aging. However, the specific mechanism underlying its role is still unknown.

The expression and localization of FMOs in the brain

The mRNAs of mammalian FMO isoforms can be detected in different organs, including the liver, kidney, lung, and brain.³⁰ FMOs are active in human, rat, mouse, rabbit, hamster, and guinea pig brains.^{31–37} Here we mainly review FMO activity in mouse and human brains.

Mouse brain

In an adult mouse brain, *FMO1* and 5 are the most abundant FMOs, as detected using isoform-specific antisense RNA probes.³⁰ *FMO1* mRNA transcripts are observed in neurons of the cerebrum and the choroid plexus while *FMO5* mRNA transcripts are only detected in neurons of the cerebrum. FMO expression in astrocytes remains controversial. Janmohamed *et al.*³⁰ reported no detectable FMO activity *in vivo*, while Di Monte *et al.*³⁸ detected FMO activity in primary cultures of mouse astrocytes.

In the neonatal brain, the most abundant FMO mRNA transcripts are *FMO1*, and their level drops by approximately 80% at 8 weeks of age. The levels of *FMO5* mRNA transcripts are 70% of *FMO1* in neonates and are similar to that of *FMO1* in 3-, 5- and 8-week-old mouse brains. *FMO2*, 3, and 4 mRNA transcripts are present at relatively low levels; approximately <1 molecule/cell.

Human brain

Zhang *et al.*³⁴ examined the developmental expression of FMOs in

60 human brain samples detecting all *FMO1–5* mRNA transcripts. FMO mRNA levels in the brain were much lower than that in other tissues, about less than 1% compared with the most abundant tissues observed (i.e., *FMO1* in the kidney, *FMO2* in the lung, and *FMO3* and 5 in the liver). *FMO1* is the only subtype to be down-regulated in adult human brains, while the amounts of other FMO mRNA transcripts in human brains remain similar among different age groups. Few studies have reported the expression of FMOs in human brains. Cashman *et al.*³⁹ found that *FMO3* was selectively expressed in the substantia nigra of human brains by immunohistochemistry.

Endogenous substances and xenobiotics oxygenated by FMOs in the brain

Endogenous substances

FMO catalyzes the N- and S-oxygenation of several endogenous substances, including phenethylamine, tyramine, amphetamine, and trimethylamine that can be converted by FMO in the brain with clinical significance.⁴⁰ S-oxygenation of hypotaurine by FMO1 contributes to the production of taurine in the brain, which possesses neurotransmitter, antioxidant, and anti-inflammatory functions.⁴¹

Xenobiotics

FMO oxidizes particular xenobiotics in the brain. Nicotine, which is abundant in tobacco smoke and can diminish oxidative stress and neuroinflammation in the brain, is hydroxylated by CYP2A6 and undergoes glucuronidation by UDP-glucuronosyl transferases and oxidation by FMO.^{21,42,43} Several psychoactive drugs, *e.g.* imipramine, chlorpromazine, and fluoxetine, are N- or S-oxygenated by FMO in both rat and human brains.^{31,32,44} Imipramine causes greater sedation in wild-type animals compared with *FMO1*-null mice, probably because imipramine N-oxide is produced in the wild-type brain and a higher concentration of desipramine is produced in the *FMO1*-null brain.⁴⁵

A typical xenobiotic oxidized by FMO is the pro-neurotoxin, MPTP, which can lead to DA neuron degeneration and parkinsonism in humans.^{46–48} MPTP in the brain is rapidly converted to the toxic MPP⁺^{49,50} by monoamine oxidase B^{51,52} or CYP (marmoset CYP2D6 and human CYP1A2).^{47,49,53} However, MPTP can be deactivated to 4-phenyl-1,2,3,6-tetrahydropyridine (PTP) and MPTP N-oxide that is non-neurotoxic, by CYP2D6 and FMO (Fig. 1).^{53–55} The concentrations of MPP⁺ in *Suncus* brains after a single intraperitoneal administration of MPTP were markedly higher than that in rats, probably because of the lack of FMO activity in *Suncus* brains.⁵⁶ *FMO1* and 3 may contribute to this detoxification. MPTP N-oxygenation in human brain microsomes was consistently catalyzed by human *FMO1* and 3.⁵³

What are the consequences of FMO deficiency?

Genetic deficiency of FMOs has several consequences. *FMO1* deficiency promotes neuroinflammation that affects the survival of DA neurons in C57BL/6N mice.²³ Mice with *FMO1*, 2, and 4 deficiency exhibit a lean phenotype and enhanced resting energy expenditure, those with *FMO1* deficiency most likely underlying the metabolic phenotype.⁵⁷ *FMO3* is a target of insulin and knockdown of *FMO3* expression in insulin-resistant mice improves glucose tolerance.⁶ Knockdown of *FMO3* expression in the liver of low-density lipoprotein receptor-knockout mice leads to decreased

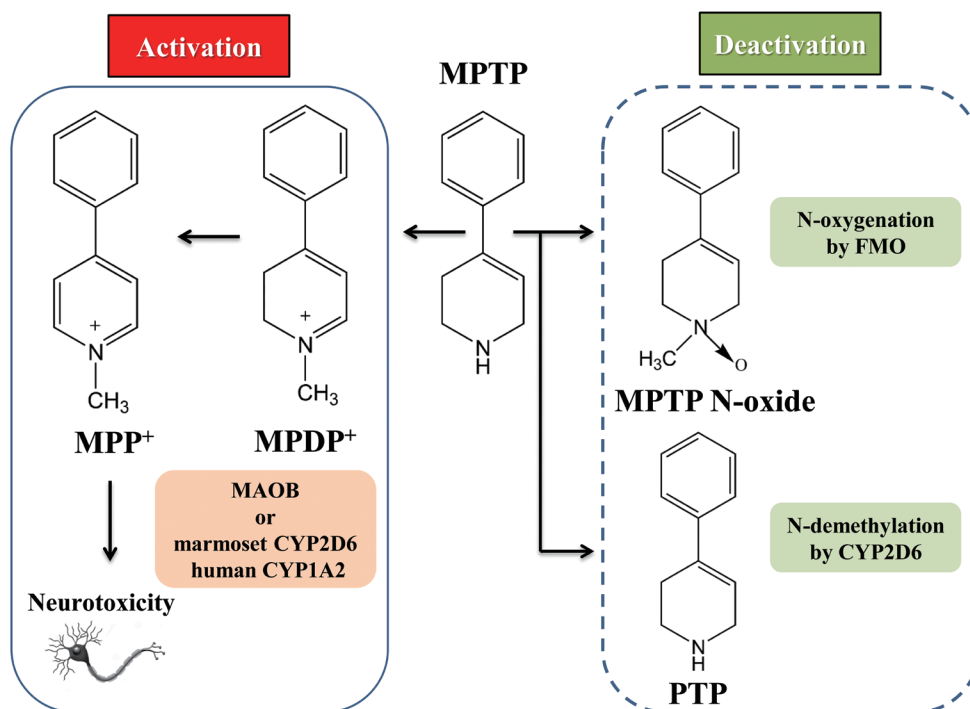


Fig. 1. Metabolic activation and deactivation of MPTP. Activation: MPTP was rapidly converted to the toxic metabolite MPP⁺ through the intermediate MPDP⁺ once in the brain mediated by monoamine oxidase B or CYP (marmoset CYP2D6 and human CYP1A2). Deactivation: MPTP N-oxygenation was efficiently mediated by FMOs in marmoset liver and brain microsomes. PTP formation was efficiently mediated by CYP2D6 in marmoset liver microsomes. FMOs, flavin-containing monooxygenases; MPTP, 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine; MPP⁺, 1-methyl-4-phenylpyridinium; PTP, 4-phenyl-1,2,3,6-tetrahydropyridine; CYP, cytochrome P450.

circulating trimethylamine N-oxide (TMAO) levels (an independent risk factor for cardiovascular disease) and atherosclerosis.^{4,5} *Fmo5*^{-/-} mice exhibit a lean phenotype and are resistant to age-related changes in glucose homeostasis compared with wild-type mice, indicating that FMO5 is a regulator of metabolic aging.⁵⁸ *Fmo5*^{-/-} mice also possess metabolic characteristics similar to those of germ-free mice, indicating that FMO5 is crucial for sensing or responding to gut bacteria.⁵⁹ However, conditional knock-down of brain FMOs has not been reported.

Further directions

The precise roles of FMOs in pathological processes remain to be determined. In-depth knowledge of FMO gene expression and protein localization and identification of substrates in the brain that are oxidized by FMOs may help in understanding the mechanisms of action of FMOs and their importance in the pathogenesis of neuronal degeneration and aging.

Conclusions

The potential involvement of FMOs in neurodegeneration and aging has been demonstrated in recent years. FMOs play important roles in metabolizing certain endogenous chemicals and xenobiotics in the brain, which participate in physiological and pathological processes. Knowledge of the expression and localization of FMOs in the brain, the endogenous chemicals and xenobiotics metabolized by FMOs, and the consequences of FMO deficiency can help us understand their involvement in neurodegeneration and aging.

Acknowledgments

We thank Dr. Shifeng Chu for his helpful advice and Jinglin Wang, Bingqing Ren, Linxi Zhang for their involvement in discussions.

Funding

This work was supported by (1) The National Natural Science Foundation of China (Grant no. 81803500), (2) The Beijing Hospitals Authority Innovation Studio of Young Staff Funding Support (code 202108), and (3) The Undergraduate Scientific Research Training Program (XSKY2022).

Conflict of interest

The authors declare no conflict of interests.

Author contributions

Writing of the original draft (BL); supervision (ZA).

References

- [1] Phillips IR, Shephard EA. Drug metabolism by flavin-containing monooxygenases of human and mouse. *Expert Opin Drug Metab Toxicol* 2017;13(2):167–181. doi:10.1080/17425255.2017.1239718, PMID:27678284.
- [2] Cashman JR. Role of flavin-containing monooxygenase in drug development. *Expert Opin Drug Metab Toxicol* 2008;4(12):1507–1521. doi:10.1517/17425250802522188, PMID:19040327.

- [3] Fennema D, Phillips IR, Shephard EA. Trimethylamine and Trimethylamine N-Oxide, a Flavin-Containing Monooxygenase 3 (FMO3)-Mediated Host-Microbiome Metabolic Axis Implicated in Health and Disease. *Drug Metab Dispos* 2016;44(11):1839–1850. doi:10.1124/dmd.116.070615, PMID:27190056.
- [4] Shih DM, Wang Z, Lee R, Meng Y, Che N, Charugundla S, *et al.* Flavin containing monooxygenase 3 exerts broad effects on glucose and lipid metabolism and atherosclerosis. *J Lipid Res* 2015;56(1):22–37. doi:10.1194/jlr.M051680, PMID:25378658.
- [5] Shih DM, Zhu W, Schugar RC, Meng Y, Jia X, Miikeda A, *et al.* Genetic Deficiency of Flavin-Containing Monooxygenase 3 (Fmo3) Protects Against Thrombosis but Has Only a Minor Effect on Plasma Lipid Levels–Brief Report. *Arterioscler Thromb Vasc Biol* 2019;39(6):1045–1054. doi:10.1161/ATVBAHA.119.312592, PMID:31070450.
- [6] Miao J, Ling AV, Manthena PV, Gearing ME, Graham MJ, Croke RM, *et al.* Flavin-containing monooxygenase 3 as a potential player in diabetes-associated atherosclerosis. *Nat Commun* 2015;6:6498. doi:10.1038/ncomms7498, PMID:25849138.
- [7] Rossner R, Kaeberlein M, Leiser SF. Flavin-containing monooxygenases in aging and disease: Emerging roles for ancient enzymes. *J Biol Chem* 2017;292(27):11138–11146. doi:10.1074/jbc.R117.779678, PMID:28515321.
- [8] Ziegler DM. Flavin-containing monooxygenases: catalytic mechanism and substrate specificities. *Drug Metab Rev* 1988;19(1):1–32. doi:10.3109/03602538809049617, PMID:3293953.
- [9] Poulsen LL, Ziegler DM. Multisubstrate flavin-containing monooxygenases: applications of mechanism to specificity. *Chem Biol Interact* 1995;96(1):57–73. doi:10.1016/0009-2797(94)03583-t, PMID:7720105.
- [10] Phillips IR, Dolphin CT, Clair P, Hadley MR, Hutt AJ, McCombie RR, *et al.* The molecular biology of the flavin-containing monooxygenases of man. *Chem Biol Interact* 1995;96(1):17–32. doi:10.1016/0009-2797(94)03580-2, PMID:7720101.
- [11] Hernandez D, Janmohamed A, Chandan P, Phillips IR, Shephard EA. Organization and evolution of the flavin-containing monooxygenase genes of human and mouse: identification of novel gene and pseudogene clusters. *Pharmacogenetics* 2004;14(2):117–130. doi:10.1097/00008571-200402000-00006, PMID:15077013.
- [12] Cereda C, Gabanti E, Corato M, de Silvestri A, Alimonti D, Cova E, *et al.* Increased incidence of FMO1 gene single nucleotide polymorphisms in sporadic amyotrophic lateral sclerosis. *Amyotroph Lateral Scler* 2006;7(4):227–234. doi:10.1080/17482960600864413, PMID:17127561.
- [13] Krueger SK, Williams DE. Mammalian flavin-containing monooxygenases: structure/function, genetic polymorphisms and role in drug metabolism. *Pharmacol Ther* 2005;106(3):357–387. doi:10.1016/j.pharmthera.2005.01.001, PMID:15922018.
- [14] Cashman JR. Structural and catalytic properties of the mammalian flavin-containing monooxygenase. *Chem Res Toxicol* 1995;8(2):166–181. doi:10.1021/tx00044a001, PMID:7766799.
- [15] Treacy EP, Akerman BR, Chow LM, Youil R, Bibeau C, Lin J, *et al.* Mutations of the flavin-containing monooxygenase gene (FMO3) cause trimethylaminuria, a defect in detoxication. *Hum Mol Genet* 1998;7(5):839–845. doi:10.1093/hmg/7.5.839, PMID:9536088.
- [16] Cashman JR, Camp K, Fakharzadeh SS, Fennessey PV, Hines RN, Mamer OA, *et al.* Biochemical and clinical aspects of the human flavin-containing monooxygenase form 3 (FMO3) related to trimethylaminuria. *Curr Drug Metab* 2003;4(2):151–170. doi:10.2174/1389200033489505, PMID:12678693.
- [17] Malaspina A, Kaushik N, de Belle Roche J. Differential expression of 14 genes in amyotrophic lateral sclerosis spinal cord detected using gridded cDNA arrays. *J Neurochem* 2001;77(1):132–145. doi:10.1046/j.1471-4159.2001.t01-1-00231.x, PMID:11279269.
- [18] Gagliardi S, Abel K, Bianchi M, Milani P, Bernuzzi S, Corato M, *et al.* Regulation of FMO and PON detoxication systems in ALS human tissues. *Neurotox Res* 2013;23(4):370–377. doi:10.1007/s12640-012-9356-1, PMID:23073612.
- [19] Gagliardi S, Oglioni P, Davin A, Corato M, Cova E, Abel K, *et al.* Flavin-containing monooxygenase mRNA levels are up-regulated in ALS brain areas in SOD1-mutant mice. *Neurotox Res* 2011;20(2):150–158. doi:10.1007/s12640-010-9230-y, PMID:21082301.
- [20] Barreto GE, Iarkov A, Moran VE. Beneficial effects of nicotine, cotinine and its metabolites as potential agents for Parkinson's disease. *Front Aging Neurosci* 2014;6:340. doi:10.3389/fnagi.2014.00340, PMID:25620929.
- [21] Perez-Paramo YX, Chen G, Ashmore JH, Watson CJW, Nasrin S, Adams-Haduch J, *et al.* Nicotine-N'-Oxidation by Flavin Monooxygenase Enzymes. *Cancer Epidemiol Biomarkers Prev* 2019;28(2):311–320. doi:10.1158/1055-9965.EPI-18-0669, PMID:30381441.
- [22] Li B, Yuan Y, Zhang W, He W, Hu J, Chen N. Flavin-containing monooxygenase, a new clue of pathological proteins in the rotenone model of parkinsonism. *Neurosci Lett* 2014;566:11–16. doi:10.1016/j.neulet.2013.11.036, PMID:24440618.
- [23] Li B, Yang S, Ye J, Chu S, Chen N, An Z. Flavin-containing monooxygenase 1 deficiency promotes neuroinflammation in dopaminergic neurons in mice. *Neurosci Lett* 2021;764:136222. doi:10.1016/j.neulet.2021.136222, PMID:34500002.
- [24] Swindell WR. Genes and gene expression modules associated with caloric restriction and aging in the laboratory mouse. *BMC Genomics* 2009;10:585. doi:10.1186/1471-2164-10-585, PMID:19968875.
- [25] Steinbaugh MJ, Sun LY, Bartke A, Miller RA. Activation of genes involved in xenobiotic metabolism is a shared signature of mouse models with extended lifespan. *Am J Physiol Endocrinol Metab* 2012;303(4):E488–E495. doi:10.1152/ajpendo.00110.2012, PMID:22693205.
- [26] Schirra HJ, Anderson CG, Wilson WJ, Kerr L, Craik DJ, Waters MJ, *et al.* Altered metabolism of growth hormone receptor mutant mice: a combined NMR metabolomics and microarray study. *PLoS One* 2008;3(7):e2764. doi:10.1371/journal.pone.0002764, PMID:18648510.
- [27] Swindell WR. Gene expression profiling of long-lived dwarf mice: longevity-associated genes and relationships with diet, gender and aging. *BMC Genomics* 2007;8:353. doi:10.1186/1471-2164-8-353, PMID:17915019.
- [28] Guo D, Shen Y, Li W, Li Q, Miao Y, Zhong Y. Upregulation of flavin-containing monooxygenase 3 mimics calorie restriction to retard liver aging by inducing autophagy. *Aging (Albany NY)* 2020;12(1):931–944. doi:10.18632/aging.102666, PMID:31927537.
- [29] Bhagwat SV, Bhamre S, Boyd MR, Ravindranath V. Cerebral metabolism of imipramine and a purified flavin-containing monooxygenase from human brain. *Neuropsychopharmacology* 1996;15(2):133–142. doi:10.1016/0893-133X(95)00175-D, PMID:8840349.
- [30] Janmohamed A, Hernandez D, Phillips IR, Shephard EA. Cell-, tissue-, sex- and developmental stage-specific expression of mouse flavin-containing monooxygenases (Fmos). *Biochem Pharmacol* 2004;68(1):73–83. doi:10.1016/j.bcp.2004.02.036, PMID:15183119.
- [31] Bhamre S, Bhagwat SV, Shankar SK, Williams DE, Ravindranath V. Cerebral flavin-containing monooxygenase-mediated metabolism of antidepressants in brain: immunochemical properties and immunocytochemical localization. *J Pharmacol Exp Ther* 1993;267(1):555–559. PMID:8229786.
- [32] Bhamre S, Bhagwat SV, Shankar SK, Boyd MR, Ravindranath V. Flavin-containing monooxygenase mediated metabolism of psychoactive drugs by human brain microsomes. *Brain Res* 1995;672(1-2):276–280. doi:10.1016/0006-8993(94)01135-5, PMID:7749747.
- [33] Zane NR, Chen Y, Wang MZ, Thakker DR. Cytochrome P450 and flavin-containing monooxygenase families: age-dependent differences in expression and functional activity. *Pediatr Res* 2018;83(2):527–535. doi:10.1038/pr.2017.226, PMID:28922349.
- [34] Zhang J, Cashman JR. Quantitative analysis of FMO gene mRNA levels in human tissues. *Drug Metab Dispos* 2006;34(1):19–26. doi:10.1124/dmd.105.006171, PMID:16183778.
- [35] Bhagwat SV, Bhamre S, Boyd MR, Ravindranath V. Further characterization of rat brain flavin-containing monooxygenase. Metabolism of imipramine to its N-oxide. *Biochem Pharmacol* 1996;51(11):1469–1475. doi:10.1016/0006-2952(96)00088-3, PMID:8630088.
- [36] Blake BL, Philpot RM, Levi PE, Hodgson E. Xenobiotic biotransforming enzymes in the central nervous system: an isoform of flavin-containing monooxygenase (FMO4) is expressed in rabbit brain. *Chem Biol Interact* 1996;99(1-3):253–261. doi:10.1016/0009-2797(95)03679-2, PMID:8620573.
- [37] Kawaji A, Isobe M, Takabatake E. Differences in enzymatic properties of flavin-containing monooxygenase in brain microsomes

- of rat, mouse, hamster, guinea pig and rabbit. *Biol Pharm Bull* 1997;20(8):917–919. doi:10.1248/bpb.20.917, PMID:9300142.
- [38] Di Monte DA, Wu EY, Irwin I, Delanney LE, Langston JW. Biotransformation of 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine in primary cultures of mouse astrocytes. *J Pharmacol Exp Ther* 1991;258(2):594–600. PMID:1907660.
- [39] Cashman JR, Zhang J. Interindividual differences of human flavin-containing monooxygenase 3: genetic polymorphisms and functional variation. *Drug Metab Dispos* 2002;30(10):1043–1052. doi:10.1124/dmd.30.10.1043, PMID:12228178.
- [40] Lin J, Cashman JR. Detoxication of tyramine by the flavin-containing monooxygenase: stereoselective formation of the trans oxime. *Chem Res Toxicol* 1997;10(8):842–852. doi:10.1021/tx970030o, PMID:9282832.
- [41] Veeravalli S, Phillips IR, Freire RT, Varshavi D, Everett JR, Shephard EA. Flavin-Containing Monooxygenase 1 Catalyzes the Production of Taurine from Hypotaurine. *Drug Metab Dispos* 2020;48(5):378–385. doi:10.1124/dmd.119.089995, PMID:32156684.
- [42] Hukkanen J, Jacob P 3rd, Benowitz NL. Metabolism and disposition kinetics of nicotine. *Pharmacol Rev* 2005;57(1):79–115. doi:10.1124/pr.57.1.3, PMID:15734728.
- [43] von Weymarn LB, Brown KM, Murphy SE. Inactivation of CYP2A6 and CYP2A13 during nicotine metabolism. *J Pharmacol Exp Ther* 2006;316(1):295–303. doi:10.1124/jpet.105.091306, PMID:16188955.
- [44] Ravindranath V, Boyd MR. Xenobiotic metabolism in brain. *Drug Metab Rev* 1995;27(3):419–448. doi:10.3109/03602539508998330, PMID:8521749.
- [45] Hernandez D, Janmohamed A, Chandan P, Omar BA, Phillips IR, Shephard EA. Deletion of the mouse *Fmo1* gene results in enhanced pharmacological behavioural responses to imipramine. *Pharmacogenet Genomics* 2009;19(4):289–299. doi:10.1097/FPC.0b013e328328d507, PMID:19262426.
- [46] Langston JW, Ballard PA Jr. Parkinson's disease in a chemist working with 1-methyl-4-phenyl-1,2,5,6-tetrahydropyridine. *N Engl J Med* 1983;309(5):310. doi:10.1056/nejm198308043090511, PMID:6602944.
- [47] Ballard PA, Tetrad JW, Langston JW. Permanent human parkinsonism due to 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine (MPTP): seven cases. *Neurology* 1985;35(7):949–956. doi:10.1212/wnl.35.7.949, PMID:3874373.
- [48] Davis KD, Taub E, Houle S, Lang AE, Dostrovsky JO, Tasker RR, *et al*. Globus pallidus stimulation activates the cortical motor system during alleviation of parkinsonian symptoms. *Nat Med* 1997;3(6):671–674. doi:10.1038/nm0697-671, PMID:9176495.
- [49] Langston JW. The MPTP Story. *J Parkinsons Dis* 2017;7(s1):S11–S19. doi:10.3233/JPD-179006, PMID:28282815.
- [50] Langston JW, Irwin I, Langston EB, Forno LS. 1-Methyl-4-phenylpyridinium ion (MPP⁺): identification of a metabolite of MPTP, a toxin selective to the substantia nigra. *Neurosci Lett* 1984;48(1):87–92. doi:10.1016/0304-3940(84)90293-3, PMID:6332288.
- [51] Chiba K, Trevor A, Castagnoli N Jr. Metabolism of the neurotoxic tertiary amine, MPTP, by brain monoamine oxidase. *Biochem Biophys Res Commun* 1984;120(2):574–578. doi:10.1016/0006-291x(84)91293-2, PMID:6428396.
- [52] Heikkila RE, Manzino L, Cabbat FS, Duvoisin RC. Protection against the dopaminergic neurotoxicity of 1-methyl-4-phenyl-1,2,5,6-tetrahydropyridine by monoamine oxidase inhibitors. *Nature* 1984;311(5985):467–469. doi:10.1038/311467a0, PMID:6332989.
- [53] Uehara S, Uno Y, Inoue T, Murayama N, Shimizu M, Sasaki E, *et al*. Activation and deactivation of 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine by cytochrome P450 enzymes and flavin-containing monooxygenases in common marmosets (*Callithrix jacchus*). *Drug Metab Dispos* 2015;43(5):735–742. doi:10.1124/dmd.115.063594, PMID:25735838.
- [54] Bajpai P, Sangar MC, Singh S, Tang W, Bansal S, Chowdhury G, *et al*. Metabolism of 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine by mitochondrion-targeted cytochrome P450 2D6: implications in Parkinson disease. *J Biol Chem* 2013;288(6):4436–4451. doi:10.1074/jbc.M112.402123, PMID:23258538.
- [55] Herraiz T, Guillén H, Galisteo J. Metabolite profile resulting from the activation/inactivation of 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine and 2-methyltetrahydro- β -carboline by oxidative enzymes. *Biomed Res Int* 2013;2013:248608. doi:10.1155/2013/248608, PMID:23984327.
- [56] Mushiroda T, Ariyoshi N, Yokoi T, Takahara E, Nagata O, Kato H, *et al*. Accumulation of the 1-methyl-4-phenylpyridinium ion in suncus (*Suncus murinus*) brain: implication for flavin-containing monooxygenase activity in brain microvessels. *Chem Res Toxicol* 2001;14(2):228–232. doi:10.1021/tx0001225, PMID:11258972.
- [57] Veeravalli S, Omar BA, Houseman L, Hancock M, Gonzalez Malagon SG, Scott F, *et al*. The phenotype of a flavin-containing monooxygenase knockout mouse implicates the drug-metabolizing enzyme FMO1 as a novel regulator of energy balance. *Biochem Pharmacol* 2014;90(1):88–95. doi:10.1016/j.bcp.2014.04.007, PMID:24792439.
- [58] Gonzalez Malagon SG, Melidoni AN, Hernandez D, Omar BA, Houseman L, Veeravalli S, *et al*. The phenotype of a knockout mouse identifies flavin-containing monooxygenase 5 (FMO5) as a regulator of metabolic ageing. *Biochem Pharmacol* 2015;96(3):267–277. doi:10.1016/j.bcp.2015.05.013, PMID:26049045.
- [59] Scott F, Gonzalez Malagon SG, O'Brien BA, Fennema D, Veeravalli S, Coveney CR, *et al*. Identification of Flavin-Containing Monooxygenase 5 (FMO5) as a Regulator of Glucose Homeostasis and a Potential Sensor of Gut Bacteria. *Drug Metab Dispos* 2017;45(9):982–989. doi:10.1124/dmd.117.076612, PMID:28646079.